Ligand Conformational Bias Drives Enantioselective Modification of a Surface-Exposed Lysine on Hsp90

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ABSTRACT: Targeted covalent modification of surface-exposed lysines is challenging due to their low intrinsic reactivity and high prevalence throughout the proteome. Strategies for optimizing the rate of covalent bond formation by a reversibly bound inhibitor (k\text{inact}) typically involve increasing the reactivity of the electrophile, which increases the risk of off-target modification. Here, we employ an alternative approach for increasing k\text{inact} of a lysine-targeted covalent Hsp90 inhibitor, independent of the reversible binding affinity (K\text{i}) or the intrinsic electrophilicity. Starting with a noncovalent ligand, we appended a chiral, conformationally constrained linker, which orients an arylsulfonyl fluoride to react rapidly and enantioselectively with Lys58 on the surface of Hsp90. Biochemical experiments and high-resolution crystal structures of covalent and noncovalent ligand/Hsp90 complexes provide mechanistic insights into the role of ligand conformation in the observed enantioselectivity. Finally, we demonstrate selective covalent targeting of cellular Hsp90, which results in a prolonged heat shock response despite concomitant degradation of the covalent ligand/Hsp90 complex. Our work highlights the potential of engineering ligand conformational constraints to dramatically accelerate covalent modification of a distal, poorly nucleophilic lysine on the surface of a protein target.

INTRODUCTION

Covalent inhibitors have broad utility as drugs, cell biological tools, and chemoproteomic probes. Irreversible covalent modification results in drug-resistant residence times that match the lifetime of the targeted protein, often independent of the drug clearance rate.\textsuperscript{9,10} Additionally, covalent inhibitors can discriminate between closely related paralogs by reacting with a nonconserved nucleophilic amino acid within or near the ligand binding site.\textsuperscript{11-13} Selective modification of the intended nucleophile is determined by a two-step reaction mechanism in which reversible binding of the ligand precedes covalent modification. Both the reversible binding affinity and the rate of covalent bond formation within the initially formed noncovalent complex (k\text{inact}) contribute to the potency of covalent inhibitors.\textsuperscript{14} An obvious way to increase k\text{inact} is to augment the intrinsic reactivity of the electrophile. A downside of this approach is that it increases the likelihood of undesired off-target reactions. Optimization of covalent inhibitors therefore relies primarily on maximizing the reversible binding affinity of the noncovalent recognition element.\textsuperscript{15-17} To date, the design of rapid-acting, highly selective covalent ligands has focused on cysteine, in part because its high intrinsic reactivity allows the use of relatively unreactive electrophiles (e.g., acrylamides).\textsuperscript{12-14} Cysteine is one of the least prevalent amino acids in the proteome, however, and many ligand binding sites lack a proximal cysteine.

Lysine is more prevalent than cysteine, and its side-chain amine is therefore a potentially attractive target for covalent inhibitor design.\textsuperscript{15,16} Nevertheless, rapid and selective covalent modification of lysine is challenging. First, the side-chain amine is mostly protonated at physiological pH and has attenuated reactivity. Second, the butylamine side chain has many conformational degrees of freedom. This enhanced flexibility results in an entropic cost to covalent capture, potentially complicating the design of inhibitors with high k\text{inact}. Third, the high prevalence of lysine in the proteome increases the probability of off-target reactions. All of these challenges are exacerbated when attempting to target a surface-exposed lysine.

Arylsulfonyl fluorides, which are weakly reactive toward water and simple amines at physiological pH, have been employed as covalent lysine-targeted inhibitors of protein kinases and stabilizers of mutant transthyretin.\textsuperscript{17-19} However, in these proteins, the targeted lysines are in hydrophobic ligand binding sites, form specific hydrogen bonds with proximal amino acid side chains, and likely have perturbed pK\textsubscript{a}s.\textsuperscript{18,20}
Whether arylsulfonyl fluorides are sufficiently reactive to rapidly modify a solvated lysine on the surface of a protein remains underexplored.21,22 Here, we report the design of arylsulfonyl fluorides that selectively target Lys58 on the surface of the heat shock protein, Hsp90, a critical regulator of cellular proteostasis.23 Although cyanoethyl acylsulfonamide probes were recently developed to modify Hsp90 on Lys58, the structural and mechanistic details of this reaction were not...
explored. By contrast, our mechanistic studies reveal how chiral conformational constraints in the ligand enantioselectively increase the rate of covalent bond formation, independent of the ligand’s reversible binding affinity.

### RESULTS AND DISCUSSION

We used the crystal structure of the Hsp90 N-terminal ATPase domain (NTD) bound to the reversible inhibitor, PU-H71, as a starting point to develop electrophilic inhibitors that target Lys58. This surface-exposed lysine lies outside the ATP pocket, 10 Å from the purine core of PU-H71 (Figure 1a). Our covalent Hsp90 inhibitors are based on a modular design, including (1) a noncovalent recognition scaffold based on PU-H71, (2) an arylsulfonyl fluoride electrophile, and most critically, (3) a variable linker to orient the arylsulfonyl fluoride toward Lys58. On the basis of the inherent flexibility of both the propylamine linker in PU-H71 and the side chain of Lys58, we reasoned that an arylsulfonyl fluoride would be tolerated as a replacement for the isopropyl group, leading to the design of arylsulfonyl fluorides 1 and 2 (Figure 1b). Treatment of recombinant Hsp90 NTD (N-terminal ATPase domain) with a saturating concentration of arylsulfonyl fluoride 1 (10 μM, 1 h) resulted in the formation of a 1:1 covalent adduct, as revealed by mass spectrometry (Figure 1c). Kinetic analysis using this assay revealed $k_{\text{on}} \sim 0.025 \text{ min}^{-1}$ ($t_{1/2} \sim 30 \text{ min}$) and $K_i \sim 60 \text{nM}$ (Figure 1d and S1), with $K_i$ being similar to the published equilibrium dissociation constant of PU-H71. Mutation of Lys58 to arginine abrogated covalent modification, despite the presence of 16 other lysines in the Hsp90 NTD (Figure 1c).

To reveal the binding mode of arylsulfonyl fluoride 1 after modification of Hsp90 NTD, we crystallized the covalent complex and determined the structure at a resolution of 1.8 Å. The purine thioether of 1 superimposes perfectly with the previously determined cocrystal structure of PU-H71. In contrast to PU-H71, the propylamine linker of 1 adopts a fully extended conformation, whereas the conformation of the Lys58 side chain is slightly kinked. Continuous electron density between Lys58 and the sulfonyl group of 1 provided unambiguous evidence for a covalent bond (Figure 1e).

We next characterized the selectivity of arylsulfonyl fluoride 1 in cells. We synthesized alkyne 2 as a clickable version of 1 suitable for cellular target engagement experiments. Skbr3 cells were treated with 2 (3.3 μM, 90 min) and probe-labeled proteins were visualized by in-gel fluorescence after copper-catalyzed conjugation of TAMRA-azide (TAMRA = tetramethylrhodamine). This revealed a prominent 90-kDa band, which was strongly competed by pretreating cells with an equivalent concentration of 1 (Figure 1f).

To identify the cellular protein targets and modification sites of probe 2 in an unbiased manner, we used a hydrazine-cleavable biotin-azide reagent. Cells were treated with 2, and covalently modified proteins were enriched with streptavidin-agarose beads after click conjugation with a hydrazine-cleavable biotin-azide reagent. After on-bead trypsinization and extensive washing, probe-modified peptides were eluted via hydrazine cleavage and analyzed by LC-MS/MS (see Supporting Information for details). Assignment of MS2 fragments are indicated in red.

Figure 2. MS/MS spectra of probe 2-labeled peptides derived from Hsp90α (a) and Hsp90β/Grp94 (b). Cells were treated with probe 2, and covalently modified proteins were enriched with streptavidin-agarose beads after click conjugation with a hydrazine-cleavable biotin-azide reagent. After on-bead trypsinization and extensive washing, probe-modified peptides were eluted via hydrazine cleavage and analyzed by LC-MS/MS (see Supporting Information for details). Assigned MS2 fragments are indicated in red.
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Because Hsp90αβ were competed to a much greater extent than Grp94 (~8-fold vs ~2-fold, Figure S2), it is likely that arylsulfonyl fluoride 1 potently engages Hsp90α and Hsp90β in cells, whereas it weakly engages Grp94. In conclusion, arylsulfonyl fluorides 1 and 2 demonstrate significant Hsp90 occupancy and appear to selectively modify Lys58 in cells.

Given that the rate of Hsp90 modification by arylsulfonyl fluoride 1 is relatively slow (t1/2 ~ 30 min, Figure 1d), a key question concerns whether the sulfonyl fluoride is optimally positioned to facilitate nucleophilic attack by Lys58 within the context of the initial reversible complex. In an attempt to reduce the entropic penalty of adopting a Lys58-reactive conformation, we constrained the propylamine linker within a 4-, 5-, or 6-membered ring to give azetidine 3, pyrrolidine 4 (synthesized as a racemic mixture) and the enantiomerically pure piperidines 5 and 6 (Figure 3a). We measured Hsp90 modification rates at a compound concentration of 10 μM, i.e., 250-fold higher than the Kd of PU-H71. Assuming the different linkers do not substantially alter the reversible binding affinity, this simple measurement should reveal the relative rates of covalent bond formation between Lys58 and reversibly bound 1 and 3–6 (i.e., relative kinact values). Strikingly, sulfonyl fluoride 5, which has the (S)-methylpiperidine linker, exhibited the fastest modification rate (t1/2 ~ 5 min), while its enantiomer 6 was the slowest compound tested (t1/2 ~ 200 min) (Figure 3b, Table S3).

To confirm that the observed modification rates primarily reflect differences in kinact and not the reversible binding affinity, we determined the relative binding affinities of 1 and 3–6 using the K58R mutant of Hsp90 NTD, which is resistant to covalent modification. Using an established fluorescence polarization assay with FITC-geldanamycin as the fluorescent ligand,27 we found that compounds 1 and 3–6 compete with FITC-geldanamycin for binding to Hsp90 with similar IC50 values (Figure 3c, Table S3). These data suggest that compounds 1 and 3–6 bind Hsp90 with similar affinities and that the observed modification rates at 10 μM are maximal, i.e., they correspond to kinact. We also measured the concentration- and time-dependent modification of Hsp90 by enantiomers 5 and 6 (Figure S3 and S4), which revealed kkinact values of 0.13 min−1 and 0.004 min−1, respectively (32-fold), and K values of 164 nM and 378 nM, respectively (2.3-fold). Together, these data indicate that the increased rate of Hsp90 modification by 5 is driven primarily by kkinact. Given that the intrinsic reactivities of sulfonyl fluorides 5 and 6 are identical, we hypothesized that the (S)-methylpiperidine linker of 5, but not the (R)-methylpiperidine linker of 6, specifically orients the arylsulfonyl fluoride toward Lys58 on the surface of Hsp90.

We used X-ray crystallography to provide structural insights into the dramatic difference in Hsp90 modification rates exhibited by the enantiomers 5 and 6. As expected, the structure of 5 bound to Hsp90 (1.8 Å resolution) revealed continuous electron density between the sulfonyl group and Lys58, consistent with covalent bond formation (Figure 4a). We next solved the structure of the noncovalent complex of 5 bound to the K58R mutant of Hsp90 (1.65 Å resolution). Although the Arg58 side chain is disordered in this structure,

Table S1. Hydrazine-mediated cleavage of the immobilized biotin linker, followed by mass spectrometry analysis, led to the identification of only two probe-modified peptides in all three biological replicates. One peptide derives from Hsp90α (aa 47–60), whereas the amino acid sequence of the second peptide matches both Hsp90β (aa 42–55) and the related paralog Grp94 (aa 103–116) (Figure 2a,b). The MS2 spectra provided unambiguous evidence for covalent modification of the intended lysine (Lys58 based on Hsp90 numbering). Although the Arg58 side chain is disordered in this structure, the Arg58 side chain is disordered in this structure, the Arg58 side chain is disordered in this structure.
of the position it occupies after reacting with Lys58, as observed in the structure of the covalent complex. Strikingly, the (R)-methylpiperidinyl linker in 6 adopts the opposite orientation upon binding to Hsp90. The critical dihedral between the methylene substituent and the piperidine chiral center of 6 (+164°) is equal in magnitude but opposite in sign to that of 5 (−177°), such that the arylsulfonyl fluoride in 6 projects away from Lys58Arg and packs against the side chain of Ile110 (Figure 4c and SS). Although this dihedral likely represents a relatively unstrained conformation, we speculate that attractive and repulsive interactions with Hsp90 (and surrounding water molecules) contribute to the conformational preferences observed in the noncovalent structures of both 5 and 6. We conclude that upon binding to Hsp90, 5 adopts a preferred orientation that places the sulfonyl fluoride within striking distance of Lys58, giving rise to the observed 40-fold enhancement in $k_{\text{on}}$ relative to 6 (Figure 3b).

We next compared the effects of 5 and 6 in Skbr3 cells. Compound 5 exhibited concentration- and time-dependent occupancy of endogenous cellular Hsp90, as revealed by reduced Hsp90 labeling by the competitive clickable probe 2 (Figure 5a and 5b). We inferred from these experiments that cellular Hsp90 was covalently modified by 5 (but not 6), because (1) the clickable probe 2 was added to cell lysates after extensively washing the cells to remove excess 5 and 6, and (2) treatment of cells with the enantiomer 6 failed to prevent labeling by probe 2 under these conditions. Continuous treatment of Skbr3 cells with compounds 5, 6, and PU-H71 (3.3 μM, 6 h) activated the heat shock response (as shown by increased Hsp70 levels) and downregulated known Hsp90 clients, including Her2 (amplified in Skbr3 cells) and phospho-Akt (p-S473), a downstream effector of Her2 signaling (Figure 5c). This is consistent with results from the fluorescence polarization assay showing equipotent reversible binding of both enantiomers 5 and 6 to Hsp90.

To explore the cellular phenotypes resulting from pulsed, covalent inhibition of Hsp90, we treated cells with saturating amounts of 5, 6, and PU-H71 (3.3 μM) for 90 min, followed by rigorous compound washout. Under these conditions, compound 5 exhibited nearly 100% occupancy immediately after washout, as determined by the absence of subsequent labeling by probe 2 (Figure 6a). As expected, similar treatment with 6 and PU-H71 failed to result in durable Hsp90 occupancy (indicated by ~100% labeling with probe 2), demonstrating washout of the reversibly bound inhibitors. Surprisingly, levels of the covalent Hsp90-5 complex decreased over time ($t_{1/2} \sim 12$ h), as shown by the time-dependent increase in Hsp90 labeling with probe 2 (added to cell lysates). Western blotting revealed that levels of total Hsp90 remained constant throughout the washout experiment. Together, these data suggest that 5-modified Hsp90 is degraded and replaced by newly synthesized, unoccupied Hsp90 (see Figure 7 below). Despite the steady decrease in 5-modified Hsp90 (~20% occupancy after 24 h), we observed a sustained heat shock response in cells that were transiently exposed to 5—but not 6 or PU-H71—as revealed by elevated levels of Hsp70 for at least 24 h post-washout (Figure 6a). Similarly, transient exposure of cells to 5 (and to a lesser extent, 6 and PU-H71) resulted in downregulation of Her2 and phospho-Akt (Figure 6b). However, in contrast to Hsp70 induction, Her2 and phospho-Akt recovered nearly to baseline levels within 24 h post-washout. Hence, the rate at which cells recover from pulsed, covalent inhibition of Hsp90 depends on the cellular

Figure 4. Cocrystal structures of 5 and 6 bound to Hsp90. (a) 1.8-Å resolution crystal structure of 5 covalently bound to Lys58 Hsp90 NTD. Electron density ($2F_o-F_i$) is shown at a contour level of 1σ. (b) Overlay of (a) with the cocrystal structure of 5 bound reversibly to K58R Hsp90 NTD. (c) Overlay of cocrystal structures of 5 and 6 reversibly bound to K58R Hsp90 NTD.

the conformation of the (S)-methylpiperidinyl linker of 5—in particular, the dihedral between the methylene substituent and the piperidine chiral center—is nearly identical in the covalent and noncovalent complexes (Figure 4b and SS). This arrangement places the electrophilic sulfur atom within ~3 Å

Figure 5. Determination of covalent Hsp90 occupancy. (a) Western blot showing concentration-dependent covalent Hsp90 occupancy in Skbr3 cells. Starred lanes represent total cellular Hsp90. (b) Western blot showing receptor occupancy that elicits Her2 signaling. The "after washout" lane was added as a control to show that the inactivated Hsp90-5 complex still occupied cellular Hsp90. (c) Western blot showing downregulation of phospho-Akt, a downstream effector of Her2 signaling. The "after washout" lane was added as a control to show that the inactivated Hsp90-5 complex still occupied cellular Hsp90.
process, with Her2 resynthesis and activation occurring at a faster rate than Hsp70 downregulation.

The apparent decrease in Hsp90 occupancy by 5—inferr
d by monitoring the time-dependent increase in Hsp90 labeling by probe 2 (Figure 6a)—could be explained by Hsp90 resynthesis during the washout period, degradation of 5-modified Hsp90, or both. To determine the fate of irreversibly inhibited Hsp90, we treated intact cells with clickable probe 2 (3.3 μM, 90 min) and monitored the covalent Hsp90-2 adduct by in-gel fluorescence after compound washout and rhodamine-azide conjugation in cell lysates. The fluorescent signal from the covalent Hsp90-2 adduct decreased steadily over 24 h, with a half-life of 6−12 h (Figure 7a). Total Hsp90 levels did not change during this time period, likely due to resynthesis of Hsp90 concomitant with degradation of the covalent Hsp90-2 adduct. In contrast to the covalent Hsp90-2 adduct, turnover of unmodified Hsp90 was negligible (Figure 7b), consistent with previous measurements of Hsp90 turnover rates in unperturbed cells (t1/2 > 36 h).28,29 We conclude that covalent modification by compounds 2 and 5 promotes degradation of Hsp90. Further studies are required to elucidate the cellular machinery that mediates this response.

CONCLUSIONS AND PERSPECTIVE

Starting with a reversible, ATP-competitive inhibitor, we designed a series of arylsulfonyl fluorides to irreversibly trap a distal, surface-exposed lysine. Although optimization of irreversible covalent inhibitors (as determined by comparing k_{inact}/K_i values) typically involves maximizing the reversible binding affinity (minimizing K_i), we pursued an alternative strategy. Here, we sought to maximize the rate of covalent bond formation within the noncovalent complex (k_{inact}) by biasing the orientation of the arylsulfonyl fluoride toward the ε-amine (proximity-induced rate acceleration). Importantly, we were able to increase k_{inact} without increasing the intrinsic reactivity of the electrophile. Our best covalent Hsp90 inhibitor, arylsulfonyl fluoride 5, employs a chiral, conformationally constrained linker to span the 10-Å distance between the purine noncovalent recognition element and the sulfonyl fluoride. We observed striking enantiodiscrimination at the level of k_{inact}, as revealed by biochemical and structural studies. Enantioselectivity was also observed in cells, such that after compound washout, only the (S)-enantiomer produced sustained covalent inactivation of endogenous Hsp90. Together, these experiments strongly support our mechanistic interpretation that, upon binding to Hsp90, the (S)-methylpiperidine 5 is poised to undergo nucleophilic attack by Lys58, whereas the dominant conformation adopted by

Figure 5. Covalent and noncovalent inhibition of Hsp90 in cells. (a) Skbr3 cells were treated with the indicated concentrations of 5 and 6 for 90 min at 37 °C. After washing the cells, lysates were prepared and treated with probe 2 (3.3 μM, 60 min, 37 °C), followed by click conjugation with TAMRA-azide and in-gel fluorescence analysis. Normalized TAMRA fluorescence intensity (90-kDa band) was plotted. The ∼100 kDa band above Hsp90 is nonspecific and was only observed when cell lysates were treated with probe 2 (as opposed to intact cells, see Figure 1f). (b) Skbr3 cells were treated with 5 and 6 (3.3 μM, 37 °C). At the indicated time points, cell lysates were prepared, treated with probe 2, and analyzed as described above. (c). Skbr3 cells were treated with 5, 6, and PU-H71 for 6 h (3.3 μM, 37 °C). Levels of Her2, total Akt, phospho-S473 Akt, Hsp70, and Hsp90 were analyzed by Western blotting.
(R)-methylpiperidine 6 places the sulfonyl fluoride far away from Lys58.

Recently, the Hamachi group reported a complementary approach for covalently targeting Hsp90, in which an electrophilic N-cyanoethyl acylsulfonamide was employed to acylate Lys58. The cyanoethyl acylsulfonamide and arylsulfonyl fluoride Hsp90 inhibitors have similar $k_{\text{inact}}$ values, both of which are within the range exhibited by cysteine-targeted covalent drugs. A major difference between the two electrophiles concerns the chemical nature of the leaving groups: fluoride vs N-cyanoethyl-N-arylsulfonamide, with the latter being more lipophilic and sterically demanding. Overall, these studies indicate that arylsulfonyl fluorides and cyanoethyl acylsulfonamides are complementary electrophiles capable of efficiently targeting a surface-exposed lysine.

Our work complements other notable studies in which chiral ligands have been shown to covalently modify their protein targets with high levels of enantioselectivity. We anticipate that the approach of targeting a surface lysine with an arylsulfonyl fluoride oriented by a chiral, conformationally constrained linker can be applied to myriad targets that lack either a reactive cysteine or a pK$_a$-perturbed lysine within the ligand binding pocket. By analyzing protein-bound small molecules in the Protein Data Bank, we estimate that the vast majority of ligandable sites are within 10 Å of a lysine ε-amine. Looking forward, we envision the use of advanced computational methods to aid in the design of chiral, conformationally constrained linkers that maximize the rate of covalent bond formation.

**ASSOCIATED CONTENT**

*Supporting Information*

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/jacs.9b09684.

Supplemental figures and tables, detailed experimental methods including synthesis and characterization of new
compounds, X-ray data collection and refinement statistics (PDF)

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Notes

The authors declare the following competing interest(s): J.T. is a cofounder and shareholder of Global Blood Therapeutics, Principia Biopharma, Kezar Life Sciences, and Cedilla Therapeutics.

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